

BIBI: Bayesian inference of breed composition

C. A. Martínez¹  | K. Khare² | M. A. Elzo¹

¹Department of Animal Sciences,
University of Florida, Gainesville, FL,
USA

²Department of Statistics, University of
Florida, Gainesville, FL, USA

Correspondence

C. A. Martínez, Department of Animal
Sciences, University of Florida,
Gainesville, FL, USA.
Email: carlosmn@ufl.edu

Summary

The aim of this paper was to develop statistical models to estimate individual breed composition based on the previously proposed idea of regressing discrete random variables corresponding to counts of reference alleles of biallelic molecular markers located across the genome on the allele frequencies of each marker in the pure (base) breeds. Some of the existing regression-based methods do not guarantee that estimators of breed composition will lie in the appropriate parameter space, and none of them account for uncertainty about allele frequencies in the pure breeds, that is, uncertainty about the design matrix. To overcome these limitations, we proposed two Bayesian generalized linear models. For each individual, both models assume that the counts of the reference allele at each marker locus follow independent Binomial distributions, use the logit link and pose a Dirichlet prior over the vector of regression coefficients (which corresponds to breed composition). This prior guarantees that point estimators of breed composition such as the posterior mean pertain to the appropriate space. The difference between these models is that model termed BIBI does not account for uncertainty about the design matrix, while model termed BIBI2 accounts for such an uncertainty by assigning independent Beta priors to the entries of this matrix. We implemented these models in a data set from the University of Florida's multibreed Angus-Brahman population. Posterior means were used as point estimators of breed composition. In addition, the ordinary least squares estimator proposed by Kuehn et al. (2011) (OLSK) was also computed. BIBI and BIBI2 estimated breed composition more accurately than OLSK, and BIBI2 had a 7.69% improvement in accuracy as compared to BIBI.

KEYWORDS

Bayes estimators, generalized linear models, genomic data, individual breed composition

1 | INTRODUCTION

Due to the benefits of heterosis effects (Dickerson, 1973), cross-breeding is commonly implemented in breeding programmes of many livestock and plants production systems. For example, in bovine production systems (dairy and beef) in tropical and subtropical regions of the world, most of the animals are cross-bred (Burrow & Prayaga, 2004; Elzo & Famula, 1985). In animal or plants production systems using hybrid individuals, knowledge of breed composition

is useful for different reasons. For instance, heterozygosity is computed using it, certain management decisions (e.g., choosing a germplasm for a given environment) are made taking it into account and it is necessary when designing cross-breeding programmes (Frkonja, Gredler, Schnyder, Curik, & Solkner, 2012; Huang, Bates, Ernst, Fix, & Steibel, 2014; Kuehn et al., 2011). In addition, in several animal species, the commercial value of an individual is largely determined by its breed composition, and determining purity plays an important role when taking breeding

decisions (Huang et al., 2014; Funkhouser et al., 2017). Furthermore, in genetic evaluation of multibreed populations, breed composition of individuals is required because this parameter and functions of it are used as explanatory variables (Elzo & Famula, 1985). Finally, knowledge of breed composition may play an important role in certain analyses such as genome-wide association (Chiang et al., 2010; Kuehn et al., 2011).

In many populations, pedigree or ancestral breed composition data are incomplete or do not exist, and the lack of any of these sources of information prevents the traditional estimation of individual breed composition. Alternatively, molecular markers can be used to estimate breed composition. This idea has been used to estimate breed composition in some species; for instance, Parker et al. (2004) used microsatellites to estimate breed composition in dogs. Hereinafter, the traditional estimator based on pedigree and ancestral breed composition records will be referred to as the pedigree-based estimator. For (not necessarily related) individuals whose parents have the same breed composition, this estimator yields the same estimate. Henceforth, groups of individuals having the same pedigree-based breed composition estimate will be denoted as breed groups. It is known that for cross-bred individuals different from F1's, the actual breed composition varies from one individual to another within the same breed group due to crossing over and chromosomal assortment taking place during meiosis, and as a consequence, the pedigree-based breed composition could be far from the actual one. Thus, the use of genomic information helps in solving this problem and permits to obtain estimates of individual breed composition in the absence of pedigree or ancestral breed composition information (Chiang et al., 2010; Frkonja et al., 2012; Gorbach et al., 2010).

The availability of genotypes for thousands of SNP markers located across the whole genome permits to estimate breed composition using genomic data. Because it is very unlikely that certain marker alleles exist only in a given breed (private alleles), the frequencies of the reference marker alleles in each base breed have been used (Chiang et al., 2010; Kuehn et al., 2011) to estimate breed composition. Kuehn et al. (2011) adapted the linear regression approach of Chiang et al. (2010) to estimate breed composition based on SNP genotypes using least squares. This methodology was used by Huang et al. (2014) to estimate breed composition in pig populations in the United States. A limitation of this approach is that the estimators are not constrained to fall in the appropriate parameter space, and consequently, out-of-range estimated breed compositions can be obtained. When using the least squares method, this problem can be overcome by carrying out a linearly constrained quadratic optimization which guarantees the solution to be in the desired space.

Funkhouser et al. (2017) used this approach to estimate breed composition in the same swine populations studied by Huang et al. (2014).

A feature of this regression problem is that frequencies of reference marker alleles in base breeds are not observable, and consequently, they have to be estimated. Statistical methods taking uncertainty about these estimates into account have not been developed yet. Alternatively, a Bayesian estimation method permits to obtain estimates in the correct space without using constrained optimization procedures using prior distributions having the appropriate support set. In addition, Bayesian methods provide a coherent way to take into account uncertainty about allele frequencies in base breeds.

Thus, the objective of this study was to develop statistical methods incorporating genomic data (genotypes for SNP markers located across the whole genome) and accounting for uncertainty about allele frequencies in base breeds to estimate individual breed composition using a Bayesian approach.

2 | MATERIALS AND METHODS

2.1 | Modelling approaches

The idea of estimating breed composition using a regression model whose design matrix is built using reference marker allele frequencies in base breeds (Chiang et al., 2010; Kuehn et al., 2011) is adopted here.

Let B be the total number of base breeds, the objective is to estimate the fraction of each one of them in every individual. For individual i , $i = 1, 2, \dots, n$, the fraction of breed j is denoted as \mathbb{b}_{ij} , $j = 1, 2, \dots, B$. The set of fractions of each breed for every individual $\mathbb{b}_i = (\mathbb{b}_{i1}, \mathbb{b}_{i2}, \dots, \mathbb{b}_{iB})$ corresponds to its breed composition; therefore, the breed composition of an individual is a B -dimensional vector. Notice that the parameter space (the set where the vector of breed composition takes values on) corresponds to a set of B non-negative real numbers such that their sum equals one (this sort of set is known as a $(B - 1)$ -dimensional simplex). Hereinafter, this set will be denoted as Ω .

Because of the restriction $\sum_{j=1}^B \mathbb{b}_{ij} = 1, \forall i = 1, 2, \dots, n$, only $B - 1$ base breeds need to be considered in the analysis. The model corresponds to a Bayesian generalized linear model (GLM) in which the stochastic component is specified as follows. Let y_i be the vector containing the number of copies of the reference allele for every marker in individual i . Because this study focuses on diploid individuals, for individual i , it is assumed that $y_{ij} \sim \text{Binomial}(2, p_{ij})$, $0 \leq j \leq m_i$, where m_i is the number of available marker genotypes for individual i . Therefore, under the assumption of linkage equilibrium, the likelihood for individual i corresponds to the product of m_i Binomial probability mass

functions. On the other hand, the canonical link (i.e., the logit link) is used; therefore, the systematic component is

$$\boldsymbol{\eta}_i = \{\eta_{ij}\}_{m_i \times 1} = \log\left(\frac{p_{ij}}{1-p_{ij}}\right) = \mathbf{x}_j \boldsymbol{\beta}_i, \quad (1)$$

where $\boldsymbol{\beta}_i$ is an unknown vector containing the regression coefficients for $B-1$ breeds which are interpreted as the fraction of each breed in individual i , X is a matrix containing the frequencies of the reference alleles in these $B-1$ base breeds and \mathbf{x}_j is its j th row. Notice that \mathbf{x}_j is common to all individuals genotyped for the j th marker locus.

Under the assumption of independence of individual genotypes, the likelihood for the complete population is

$$\begin{aligned} L(\boldsymbol{\beta}; \mathbf{y}) &= f(\mathbf{y}|\boldsymbol{\beta}) = \prod_{i=1}^n f(\mathbf{y}_i|\boldsymbol{\beta}_i) \\ &= \prod_{i=1}^n \prod_{j=1}^{m_i} f(y_{ij}|p_{ij}) \propto \prod_{i=1}^n \frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i \rangle)}{g(\boldsymbol{\beta}_i, X)} \end{aligned} \quad (2)$$

where $\mathbf{y} := (\mathbf{y}'_1, \dots, \mathbf{y}'_n)'$, $\boldsymbol{\beta} := (\boldsymbol{\beta}'_1, \dots, \boldsymbol{\beta}'_n)'$, $\mathbf{p} := (\mathbf{p}'_1, \dots, \mathbf{p}'_n)'$, $\mathbf{p}_i := (p_{i1}, \dots, p_{im_i})'$, $1 \leq i \leq n$, operator $\langle \cdot, \cdot \rangle$ represents the dot product, $\mathbf{S}_i := \sum_{j \in M_i} y_{ij} \mathbf{x}_j$, $g(\boldsymbol{\beta}_i, X) = \prod_{j \in M_i} (1 + \exp(\langle \mathbf{x}_j, \boldsymbol{\beta}_i \rangle))^2$ and M_i is the set of marker loci for which individual i has been genotyped, the size of this set is m_i . A natural choice for the prior of $\boldsymbol{\beta}_i$ is a Dirichlet ($\boldsymbol{\alpha}_i$) distribution because its support set corresponds to the appropriate parameter space; thus, $\boldsymbol{\beta}_i \stackrel{\text{ind}}{\sim} \text{Dirichlet}(\boldsymbol{\alpha}_i)$, $1 \leq i \leq n$, $\boldsymbol{\alpha}_i = (\alpha_{i1}, \dots, \alpha_{iB})$, hence:

$$\pi(\boldsymbol{\beta}_i) \propto \prod_{k=1}^{B-1} \beta_{ik}^{\alpha_{ik}-1} \left(1 - \sum_{k=1}^{B-1} \beta_{ik}\right)^{\alpha_{iB}-1}.$$

Hereinafter, this model will be referred to as BIBI.

2.1.1 | Remark 1

Notice that this estimation problem has the following features. Unlike other regression models, the design matrix is not observable; therefore, it has to be estimated. The usual approach uses estimates obtained using pure-bred animals from a reference population (Chiang et al., 2010; Huang et al., 2014; Kuehn et al., 2011). Another feature of the problem is that for certain individuals, the true value of the parameter $\boldsymbol{\beta}_i$ is known. These are pure-bred and F1 individuals. For example, suppose that the model is parameterized in terms of breeds 1, 2, ..., $B-1$, then, if individual i is a purebred of breed k , $1 \leq k \leq B-1$, then, $\boldsymbol{\beta}_i = (0, \dots, 0, 1, 0, \dots, 0)'$ where scalar 1 is located in the k th entry of $\boldsymbol{\beta}_i$, if individual i is a purebred of breed B then $\boldsymbol{\beta}_i = \mathbf{0}_{(B-1) \times 1}$, if individual i is an F1 resulting from breeds k and k' , $1 \leq k < k' \leq B-1$, then the only non-null entries of $\boldsymbol{\beta}_i$ are k and k' each being equal to $1/2$, and so on.

Under a quadratic error loss, the Bayes estimator (i.e., the estimator minimizing the Bayesian risk) is the posterior mean (Lehmann and Casella, 1998); this is the point estimator used here. For $1 \leq i \leq n$, the posterior mean has the form

$$\begin{aligned} \hat{\boldsymbol{\beta}}_i &= \{\hat{\beta}_{ij}\}_{(B-1) \times 1} \\ &= E[\boldsymbol{\beta}_i | \mathbf{y}_i, X] \\ &= \frac{\int_{\Omega} \boldsymbol{\beta}_i \prod_{k=1}^{B-1} \beta_{ik}^{\alpha_{ik}-1} (1 - \sum_{k=1}^{B-1} \beta_{ik})^{\alpha_{iB}-1} \frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i \rangle)}{g(\boldsymbol{\beta}_i, X)} d\boldsymbol{\beta}_i}{\int_{\Omega} \prod_{k=1}^{B-1} \beta_{ik}^{\alpha_{ik}-1} (1 - \sum_{k=1}^{B-1} \beta_{ik})^{\alpha_{iB}-1} \frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i \rangle)}{g(\boldsymbol{\beta}_i, X)} d\boldsymbol{\beta}_i} \end{aligned} \quad (3)$$

Therefore, the corresponding estimator of breed composition of individual i is $\hat{\boldsymbol{\beta}}_i = (\hat{\beta}_{i1}, \dots, \hat{\beta}_{i(B-1)}, 1 - \sum_{k=1}^{B-1} \hat{\beta}_{ik})$. By properties of the expected value (Casella and Berger, 2002), it follows that the estimated breed composition lies in Ω ; consequently, this method guarantees estimates in the appropriate space. Notice that this expectation can be seen as the ratio of the expectation of two functions of $\boldsymbol{\beta}_i$ taken with respect to the prior distribution, that is, with respect to a Dirichlet ($\boldsymbol{\alpha}_i$). Thus,

$$\hat{\boldsymbol{\beta}}_i = \frac{E_{\pi} \left[\boldsymbol{\beta}_i \frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i \rangle)}{g(\boldsymbol{\beta}_i, X)} \right]}{E_{\pi} \left[\frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i \rangle)}{g(\boldsymbol{\beta}_i, X)} \right]} \quad (4)$$

Consequently, if the interest is only in estimating the posterior mean, there is no need for using Markov chain Monte Carlo (MCMC) methods because this expectation can be approximated using classical Monte Carlo integration (MCI). Therefore, once N samples $\boldsymbol{\beta}_i^1, \dots, \boldsymbol{\beta}_i^N$ are drawn from a Dirichlet ($\boldsymbol{\alpha}_i$) distribution, the Monte Carlo approximation to $\hat{\boldsymbol{\beta}}_i$ is

$$\hat{\boldsymbol{\beta}}_i^{MC} = \frac{\sum_{j=1}^N \boldsymbol{\beta}_i^j \frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i^j \rangle)}{g(\boldsymbol{\beta}_i^j, X)}}{\sum_{j=1}^N \frac{\exp(\langle \mathbf{S}_i, \boldsymbol{\beta}_i^j \rangle)}{g(\boldsymbol{\beta}_i^j, X)}} \quad (5)$$

2.1.2 | Accounting for uncertainty about the design matrix

As mentioned in Remark 1, this estimation problem exhibits the feature that the design matrix is not observable. In the previous section, a model conditioned on this matrix was proposed. The use of a Bayesian approach has the advantage of permitting to account for uncertainty about matrix X in an easy way. To this end, X is given a prior distribution. Recall that x_{kj} , $1 \leq k \leq m = \max_{1 \leq i \leq n} (m_i)$, $1 \leq j \leq B-1$ corresponds to the frequency of the reference allele of marker k in base breed j , assuming independence of these random variables across breeds (i.e., independence of columns of X) and linkage equilibrium (i.e.,

independence of rows of X), independent Beta distributions can be posed over each entry of X . The other components of the model are the same; thus, the systematic component is the one shown in Equation 1, the likelihood corresponds to the one in Equation 2 and vectors $\{\beta_i\}_{i=1}^n$, are given independent Dirichlet (α) priors. Under this model, the joint posterior is as follows:

$$\begin{aligned} \pi(\beta, X|y) &\propto \prod_{i=1}^n \frac{\exp(\langle S_i, \beta_i \rangle)}{g(\beta_i, X)} \prod_{k=1}^{B-1} \beta_{ik}^{\alpha_{ik}-1} \\ &\times \left(1 - \sum_{k=1}^{B-1} \beta_{ik}\right)^{\alpha_{iB}-1} \prod_{j=1}^m \prod_{k=1}^{B-1} x_{jk}^{\alpha_{jk}-1} (1 - x_{jk})^{b_{jk}-1} \end{aligned} \quad (6)$$

From Equation 6, it follows that all the full conditionals are not standard distributions and that given β and y the rows of X are conditionally independent, but its columns are not. In this case, the dimensionality of the problem increases notably because m unknown parameters are introduced in the model. Like in the previous scenario, the posterior mean can be used as a point estimator of breed composition and it can also be computed as a ratio of expectations taken with respect to the prior distribution. Let $\theta = (\beta', \text{vec}(X)')$ and $\Theta = \Omega \times \Gamma^{m(B-1)}$ where $\text{vec}(\cdot)$ is the vec operator and $\Gamma^{m(B-1)}$ is an $m(B-1)$ -dimensional unit hypercube. Thus, the extended model accounting for uncertainty about X has parameter θ and parameter space Θ and for $1 \leq l \leq (n+m)(B-1)$:

$$\hat{\theta}_l = E[\theta_l|y] = \frac{\int_{\Theta} \theta_l L(\theta; y) \pi(\theta) d\theta}{\int_{\Theta} L(\theta; y) \pi(\theta) d\theta} = \frac{E_{\pi}[\theta_l L(\theta; y)]}{E_{\pi}[L(\theta; y)]}$$

where

$$\begin{aligned} \pi(\theta) &= \pi(\beta, X) \propto \prod_{i=1}^n \left(\prod_{k=1}^{B-1} \beta_{ik}^{\alpha_{ik}-1} \right) \left(1 - \sum_{k=1}^{B-1} \beta_{ik}\right)^{\alpha_{iB}-1} \\ &\times \prod_{j=1}^m \prod_{k=1}^{B-1} x_{jk}^{\alpha_{jk}-1} (1 - x_{jk})^{b_{jk}-1} \end{aligned}$$

and $L(\theta; y)$ is the likelihood.

In particular, the Monte Carlo approximation to the posterior mean of β_i based on a sample $\theta^1, \dots, \theta^N$ drawn from $\pi(\theta)$ is

$$\hat{\beta}_{2i}^{MC} = \frac{\sum_{j=1}^N \beta_i^j \frac{\exp(\langle S_i^j, \beta_i^j \rangle)}{g(\beta_i^j, X^j)}}{\sum_{j=1}^N \frac{\exp(\langle S_i^j, \beta_i^j \rangle)}{g(\beta_i^j, X^j)}} \quad (7)$$

where $S_i^j = \sum_{k \in M_i} y_{ik} x_k^j$. This extended version of the proposed model will be referred to as BIBI2. For the sake of clarity, a toy example showing the computation of breed composition using expressions 5 and 7 is presented in Appendix S1.

2.2 | Assessment of accuracy

Some previous studies have considered the pedigree-based breed composition estimates (Kuehn et al., 2011) or the true breed composition from simulated data (Funkhouser et al., 2017) to assess the adequacy of estimators of individual breed composition. When working with real data, using the second property of the estimation problem being considered in this study (see Remark 1), the accuracy of the point estimators proposed here, that is, the posterior mean estimated using Equation 5 (when not accounting for uncertainty about X) or Equation 7 (when accounting for uncertainty about X) can be computed using the l_1 norm of the difference between β_i and $\hat{\beta}_i$ for pure-bred and F1 individuals. Specifically, the following measure of accuracy was used:

$$\frac{1}{|V|} \sum_{i \in V} \|\hat{\beta}_i - \beta_i\|_1 \quad (8)$$

where V is the set pure-bred and F1 individuals, $|V|$ its number of elements and operator $\|\cdot\|_1$ represents the l_1 norm.

2.3 | Implementation in a multibreed Bovine population

Implementation with real data was carried out using a multibreed beef cattle population from the University of Florida's Beef Research Unit (BRU). This multibreed population was created in 1989, and it is composed of two breeds: Angus and Brahman. Individuals are mated according to a diallel design; therefore, this population contains individuals with breed composition ranging from pure-bred Angus to pure-bred Brahman. Details on this multibreed population can be found in Elzo and Wakeman (1998). Data came from a subset of 120 individuals with pedigree-based estimated Angus fraction ranging from 0 to 1 that were genotyped for SNP markers included in the GeneSeek Genomic Profiler F-250 (Neogen Corporation, Lansing, MI, USA). Only those markers in common with the Illumina BovineSNP50 array (Illumina Inc., San Diego, CA, USA) were considered. To construct the design matrix for model BIBI and to define the prior for X in model BIBI2, the reference allele frequencies in Angus and Brahman reported by Kuehn et al. (2011) were used. These values were the entries of the design matrix when fitting BIBI, whereas when fitting BIBI2, the hyperparameters were chosen in such a way that prior means matched them. Markers located in autosomes that had a minor allele frequency larger than 0.05 were considered. Then, marker loci for which all individuals were heterozygous were removed. After this editing, a total of 9906 markers were left. In addition to models BIBI and BIBI2, the model proposed by Kuehn

et al. (2011) was fitted to this data. Accuracy of estimators was assessed using Equation 8 in a set of 60 individuals known to be purebreds (Angus and Brahman) and F1. Henceforth, this set will be referred to as MAB-V. In addition to the average in Equation 8, other descriptive statistics of the norms $\|\hat{\beta}_i - \beta_i\|_1$ were also computed. Finally, Pearson correlation coefficients among the different estimators of Angus fraction were computed in the complete set of 120 individuals and in the subset of 60 non-F1 cross-bred individuals.

3 | RESULTS

According to the expression presented in Equation 8, from the three models considered here, BIBI2 had the best performance and the estimator proposed by Kuehn et al. (2011) had the worst. Moreover, estimates from BIBI2 were less variable across individuals than those obtained from the other two models (Table 1).

On the other hand, correlations between estimated Angus fractions from the four estimators considered here (those from the three regression models mentioned above and the pedigree-based estimator) were high in both the complete set of 120 individuals and the subset of 60 non-F1 cross-bred individuals (Table 2).

The smallest value was 0.92; it corresponded to the correlation between the estimates from the method proposed by Kuehn et al. (2011) and the pedigree-based estimates in

TABLE 1 Descriptive statistics of the l_1 norm of the difference between estimated and true breed compositions in the MAB-V set

Statistic	BIBI	BIBI2	KOLS ^a
Mean	0.026	0.024	0.200
Standard deviation	0.003	0.001	0.052
Minimum	0.011	0.021	0.067
Maximum	0.028	0.026	0.266

^aOrdinary least squares estimator based on the model proposed by Kuehn et al. (2011).

TABLE 2 Pearson correlation coefficients among different kinds of Angus fraction estimates in the complete set and in the subset of non-F1 cross-bred individuals^a

	BIBI	BIBI2	KOLS	Pedigree-based
BIBI	1	0.94	0.92	0.94
BIBI2	0.95	1	0.92	0.94
KOLS	0.92	0.92	1	0.92
Pedigree-based	0.95	0.95	0.92	1

^aUpper off-diagonal elements correspond to correlations in the subset of non-F1 cross-bred individuals, while lower off-diagonal elements correspond to correlations in the complete set.

the two sets, and the largest was 0.95, corresponding to the correlations between BIBI and the pedigree-based estimates and between BIBI and BIBI2 estimates in the complete set (Table 2).

4 | DISCUSSION

In this study, Bayesian GLM's to estimate breed composition in diploid species using biallelic molecular markers were developed. Specifically, these models correspond to Bayesian logistic regressions, assigning a Dirichlet prior to the vector of regression coefficients. This prior was chosen because its support set matches the parameter space of this estimation problem (Ω). In a first stage, the model was formulated conditional on the design matrix X (BIBI), which is formed using marker allele frequencies in base breeds. However, due to the fact that these frequencies are unknown, this estimation problem has as special feature the fact that the design matrix is not observable; therefore, it has to be estimated. Consequently, this model was expanded to account for uncertainty about the design matrix (BIBI2); this expanded version treats the design matrix as an unknown parameter, and independent Beta priors are assigned to each one of its entries. Thus, this model takes into account the fact that there exists uncertainty about the design matrix and incorporates it in the estimation process. The model conditional on the design matrix is simpler because for each individual, it only has B-1 parameters. The point estimator used here was the posterior mean, which according to decision theory, minimizes the Bayesian risk under the squared error loss (Lehmann and Casella, 1998). In both models, the posterior and full conditional distributions are unknown.

In a regression model, when the explanatory variables are recorded with errors, it could induce bias in the parameter estimates; therefore, there exist approaches to take the measurement error in regression variables into account (Lehmann and Casella, 1998; Rawlings, Pantula, & Dickey, 1998). In a similar spirit, in the problem addressed here, the regression variables are actually estimates, thus, they may differ from the true values. Hence, the model accounting for this fact is theoretically more rigorous and it could be expected to yield more accurate estimators of breed composition. A small increment in accuracy (as measured by the expression in Equation 8) was observed in this study (Table 1). Relative to the accuracy of method BIBI, it corresponds to an improvement of 7.69%. At this point, it needs to be mentioned that beyond the potential gains in accuracy, model BIBI2 features the following practical advantages. Firstly, when a reference population to estimate allele frequencies in base breeds is not available, BIBI2 still permits to estimate individual breed composition.

Secondly, if it is of interest, estimates of the entries of the design matrix, namely allele frequencies in base populations can be computed because these are model parameters in BIBI2. Consequently, BIBI2 can be regarded as a more versatile and theoretically rigorous model.

Bayesian estimation of breed composition has another appealing feature. In certain cases, there is interest in making probabilistic statements about breed composition; for example, to answer the question: what is the probability that the fraction of breed k in individual i is larger than t ? Models such as BIBI and BIBI2 permit to easily answer this kind of questions using the posterior distribution of β . Specifically, if Ψ is a proper subset of Ω , then

$$P(\beta \in \Psi | y) = \int_{\Psi} \pi(\beta | y) d\beta \quad (9)$$

Notice that for model BIBI2, obtaining $\pi(\beta | y)$ requires integration of the joint posterior density of β and X with respect to X and that for models BIBI and BIBI2, the integral in Equation 9 has to be calculated numerically. In addition, interval estimates of breed composition can be obtained by computing credible sets.

Regarding evaluation of accuracy of the proposed estimators, using the fact that in this estimation problem the true value of β_i is known for pure-bred and F1 individuals, it was proposed to use this set to assess the accuracy of a given estimator of breed composition. An approach in the same spirit, but considering only pure-bred individuals from some breed of interest has been used in previous studies (Frkonja et al., 2012; Huang et al., 2014; Funkhouser et al., 2017). Notice that for pure-bred and F1 individuals, the pedigree-based estimator is 100% accurate; thus, Equation 8 is useful to compare marker-based estimators because the pedigree-based estimator will always have a value of zero. A drawback of this measure is that it does not consider the whole parameter space; it only assesses the performance of different estimators at certain points of this space. Due to the biological reasons explained in the introduction, marker-based estimators could be expected to outperform the pedigree-based estimator in non-F1 cross-bred individuals because they consider individual variation in breed composition; recall that the pedigree-based estimator takes the same value for all the offspring of a given pair of individuals, whereas a marker-based estimator does not. Kuehn et al. (2011) used the pedigree-based estimates of breed composition of cross-bred individuals to assess the performance of their regression-based estimator. However, due to the disadvantages of the pedigree-based estimator mentioned above, this approach may not be entirely appropriate to assess accuracy. Simulation is a valuable tool to assess the accuracy of different estimators of breed composition. Funkhouser et al. (2017) carried out a

simulation study to assess the performance of their constrained regression method and they found correlations between estimated and true breed compositions of 0.97.

High correlations between estimates of Angus fraction from the four methods considered here were found (Table 2). This result is in agreement with results reported by Kuehn et al. (2011) and Frkonja et al. (2012). For the set of individuals analysed in this study, although correlations between estimates of Angus fraction obtained from the Bayesian estimators and the least squares estimator proposed by Kuehn et al. (2011) were high, the accuracy of the Bayesian estimators was considerably higher.

As discussed previously, marker-based estimators of individual breed composition are a useful and reliable tool to infer breed composition when pedigree and/or ancestral breed composition records are not available. However, the benefits of marker-based estimators go beyond this scenario because having a better knowledge of individual breed composition has several potential applications even when pedigree and ancestral breed compositions are known. Examples of problems in which having marker-based estimates of breed composition could have a positive impact as compared to pedigree-based estimates are selection of candidates for cross-breeding programs, creation of the so-called synthetic breeds or creation of genetic lines, estimation of heterosis effects (Dickerson, 1973), and estimation of average breed additive effect and multibreed genetic values (Elzo & Famula, 1985). As to the latter topic, the fact that for non-F1 cross-bred individuals, pedigree-based breed composition estimates could be far apart from the actual composition poses a question regarding the impact of the use of marker-based estimates on the accuracy of predicted multibreed genetic values as defined in Elzo and Famula (1985). Breed composition and functions of it are used as explanatory variables in the linear models used to predict genetic values and to estimate genetic parameters in multibreed populations (Cardoso & Tempelman, 2004; Elzo, 1994, 1996; Elzo & Famula, 1985); therefore, it could be of interest to compare the performance of a multibreed model considering marker-based estimated breed compositions with the standard model that uses pedigree-based estimates. Our methods could also be used in across-country multibreed genetic evaluation by obtaining estimates of individual breed composition in each country and using them to build explanatory variables used in models like those described in Elzo and Bradford (1985). In these models, it is important to account for heterogeneity of variance and covariance components and genetic effects across countries; therefore, using breed compositions in each country could increase the reliability of predicted breeding values. Notice that these methods could also be applied to populations where there exist genetic groups (different from breed groups) and there have been mating between

individuals from different groups. An example of this kind of genetic groups is the case of genetically distinct populations of individuals from the same breed like those performing in different environments under different selection criteria. In this scenario, instead of predicting the fraction of each breed in each individual, the model would predict the fraction of each group and this information could be implemented in genetic evaluation models.

An example of a successful application of marker-based estimation of individual breed composition is the adoption of the methods developed by Funkhouser et al. (2017) by the US National Swine Registry (NSR) to screen pure-bred Yorkshire pigs.

For the sake of simplicity, our models do not account for linkage disequilibrium or kinship. Regarding the linkage equilibrium assumption, to meet it, the set of SNP to be used could be selected on the basis of an algorithm that picks independent markers such as tag SNP. Moreover, studying the robustness of our methods to deviations from the linkage equilibrium assumption could be part of future research. Another refinement of models BIBI and BIBI2 is to consider kinship. It can be accomplished by removing the assumption of conditional independence of vectors y_1, \dots, y_n given X and β . When pedigree information is available, a likelihood based on genetic relationships can be derived using the pedigree by following a derivation similar to the one presented in Martínez, Khare, Banerjee, and Elzo (2017). A point that also opens a path for refinement of our models is to accommodate genetic relationships when estimating allele frequencies in the base breeds. For model BIBI, the method proposed by McPeck, Wu, and Ober (2004) which yields BLUE of allele frequencies using pedigree information, could be used to estimate X . On the other hand, this could be achieved through hierarchical modelling in model BIBI2.

An alternative approach to estimate breed composition is to use distance-based or model-based clustering methods that have been designed to infer population structure. Examples of software implementing this sort of methods are STRUCTURE (Pritchard, Stephens, & Donnelly, 2000), MENDEL (Lange et al., 2001) and EigenStrat (Price et al., 2006). Some of them perform a soft clustering, that is, they compute the probability that each individual pertains to each cluster (Hastie, Tibshirani, & Friedman, 2009). Because the focus of this study was on the regression approach, these alternative methods were not implemented. Moreover, previous studies have reported high degree of agreement among the estimates from some of these methods and the regression approach (Chiang et al., 2010; Kuehn et al., 2011). Notwithstanding, it is worth mentioning the following drawback of some clustering methods. They are designed to infer a “cryptic” population structure, which is useful when the underlying factors inducing

genetic heterogeneity (structure) in the population are not easily detected. In fact, they can infer the number of subpopulations (i.e., the number of clusters); however, when using them to estimate breed composition, the number of clusters is set equal to the number of breeds. Thus, for each individual what is estimated is the probability of pertaining to each one of these clusters; consequently, breed fractions are not estimated directly and clusters have to be assigned to breeds, for example, cluster 1 corresponds to breed B and cluster 2 corresponds to breed A. Usually, this assignment is performed using pure-bred individuals or individuals known to have a high frequency of certain breed. For two or three breeds, this is not too difficult, but when the number of breeds increases and pedigree-based estimates or pure-bred individuals of certain breeds are not available, assigning clusters to breeds could be difficult and this may introduce additional bias. For example, consider the problem of estimating breed composition in a cross-bred population composed of eight breeds where pure-bred individuals and pedigree are not available.

5 | FINAL REMARKS

The use of genomic data permits to estimate breed composition when pedigree or ancestral breed composition information is missing, but advantages of this kind of estimators go beyond this. As an example, consider the potential benefits of the use of marker-based estimates of breed composition in different genetics problems that were discussed here. Therefore, this approach could be useful even when pedigree and ancestral breed composition records are available. Hence, the potential applications of these estimators open a path for further research. The regression models developed here permit to easily obtain point and interval estimates of breed composition and to make useful probabilistic statements about it. Also, unlike other regression models used to estimate breed composition, model BIBI2 accounts for uncertainty about allele frequencies in base breeds and permits to estimate breed composition in the absence of reference populations to estimate them.

ACKNOWLEDGEMENTS

Funds to obtain the data used in this study came from the following research projects: Hatch Project FLA-ANS-005150, Florida Beef Council Project 00098682 and Florida Cattle Enhancement Fund Project 00125199. Authors thank Dr. Raluca Mateescu from the Department of Animal Sciences of University of Florida for providing genotypic data and Dr. Juan Pedro Steibel from the Department of Animal Science and the Department of Fisheries and Wildlife of Michigan State University for useful discussions.

C.A. Martínez thanks Fulbright Colombia and “Departamento Administrativo de Ciencia, Tecnología e Innovación” COLCIENCIAS for supporting his PhD and master’s programs at the University of Florida through a scholarship, and Bibiana Coy for her love, support and constant encouragement.

ORCID

C. A. Martínez  <http://orcid.org/0000-0003-2842-5580>

REFERENCES

- Burrow, H. M., & Prayaga, K. C. (2004). Correlate responses in productive and adaptative traits and temperament following selection for growth and heat resistance in tropical beef cattle. *Livestock Production Science*, *86*, 143–161.
- Cardoso, F. F., & Tempelman, R. J. (2004). Hierarchical Bayes multiple-breed inference with an application to genetic evaluation of a Nelore-Hereford population. *Journal of Animal Science*, *82*, 1589–1601.
- Casella, G., & Berger, R. (2002). *Statistical Inference*, 2nd ed. Pacific Grove, CA: Duxbury.
- Chiang, C. W. K., Gajdos, Z. K. Z., Korn, J. M., Kuruvilla, F. G., Butler, J. L., Hackett, R., ... Hirschhorn, J. N. (2010). Rapid assessment of genetic ancestry in populations of unknown origin by genome-wide genotyping of pooled samples. *PLoS Genetics*, *6* (3), e1000866.
- Dickerson, G. E. (1973). Inbreeding and heterosis in animals. *Proceedings of the Animal Breeding and Genetics Symposium in Honor of Dr. J.L. Lush*, 54–77.
- Elzo, M. A. (1994). Restricted maximum likelihood estimation of additive and nonadditive genetic variances and covariances in multi-breed populations. *Journal of Animal Science*, *72*, 3055–3065.
- Elzo, M. A. (1996). Unconstrained procedures for the estimation of positive definite covariance matrices using restricted maximum likelihood in multibreed populations. *Journal of Animal Science*, *74*, 317–328.
- Elzo, M. A., & Bradford, G. E. (1985). Multibreed sire evaluation across countries. *Journal of Animal Science*, *60*, 953–963.
- Elzo, M. A., & Famula, T. R. (1985). Multibreed sire evaluation within a country. *Journal of Animal Science*, *60*, 942–952.
- Elzo, M. A., & Wakeman, D. L. (1998). Covariance components and prediction for additive and nonadditive preweaning growth genetic effects in an Angus-Brahman multibreed herd. *Journal of Animal Science*, *76*, 1290–1302.
- Frkonja, A., Gredler, B., Schnyder, U., Curik, I., & Solkner, J. (2012). Prediction of breed composition in an admixed cattle population. *Animal Genetics*, *43*, 696–703.
- Funkhouser S.A., Bates R.O., Ernst C.W., Newcom D., & Steibel J.P. (2017). Estimation of genome-wide and locus-specific breed composition in pigs. *Translational Animal Science*, *1*, 36–44.
- Gorbach, D. M., Makgahlela, M. L., Reecy, J. M., Kemp, S. J., Baltenweck, I., Ouma, R., ... Rothschild, M. F. (2010). Use of SNP genotyping to determine pedigree and breed composition of dairy cattle in Kenya. *Journal of Animal Breeding and Genetics*, *127*, 348–351.
- Hastie, T., Tibshirani, R., & Friedman, J. (2009). *The elements of statistical learning. Data mining, inference, and prediction*, 2nd ed. New York, NY: Springer.
- Huang, Y., Bates, R. O., Ernst, C. W., Fix, J. S., & Steibel, J. P. (2014). Estimation of U.S. Yorkshire breed composition using genomic data. *Journal of Animal Science*, *92*, 1395–1404.
- Kuehn, L. A., Keele, J. W., Bennett, G. L., McDaniel, T. G., Smith, T. P. L., Snelling, W. M., ... Thallman, R. M. (2011). Predicting breed composition using breed frequencies of 50,000 markers from the US Meat Animal Research Center 2,000 Bull Project. *Journal of Animal Science*, *89*, 1742–1750.
- Lange, K., Cantor, R., Horvath, S., Perola, M., Sabatti, C., Sinsheimer, J., & Sobel, E. (2001). Mendel version 4.0: A complete package for the exact genetic analysis of discrete traits in pedigree and population datasets. *American Journal of Human Genetics*, *69* (Suppl), 504.
- Lehmann E.L., & Casella G. (1998). *Theory of point estimation*. Springer, New York, NY.
- Martínez, C. A., Khare, K., Banerjee, A., & Elzo, M. A. (2017). Joint genome-wide prediction in several populations accounting for randomness of genotypes: A hierarchical Bayes approach. I: Multivariate Gaussian priors for marker effects and derivation of the joint probability mass function of genotypes. *Journal of Theoretical Biology*, *417*, 8–19.
- McPeck, M. S., Wu, X., & Ober, C. (2004). Best linear unbiased allele-frequency estimation in complex pedigrees. *Biometrics*, *60*, 359–367.
- Parker, H. G., Kim, L. V., Sutter, N. B., Carlson, S., Lorentzen, T. D., Malek, T. B., ... Kruglyak, L. (2004). Genetic structure of the purebred domestic dog. *Science*, *304*, 1160–1164.
- Price, A. L., Patterson, N. J., Plenge, R. M., Weinblatt, M. E., Shadick, N. A., & Reich, D. (2006). Principal components analysis correct for stratification in genome-wide association studies. *Nature Genetics*, *38*, 904–909.
- Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using multilocus genotype data. *Genetics*, *155*, 945–959.
- Rawlings, J. O., Pantula, S. G., & Dickey, D. A. (1998). *Applied regression analysis. A research tool*, 2nd ed. New York, NY: Springer.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

How to cite this article: Martínez CA, Khare K, Elzo MA. BIBI: Bayesian inference of breed composition. *J Anim Breed Genet*. 2018;135:54–61. <https://doi.org/10.1111/jbg.12305>